

Study of Virulence Factors and Antimicrobial Susceptibility Pattern of Uropathogenic Escherichia Coli in a Tertiary Care Hospital

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ABSTRACT

Introduction: Urinary tract infections are the most commonly encountered bacterial infections both in community and health care settings. Uropathogenic Escherichia coli (UPEC) is the single most common pathogen, accounting for 70-75% of all cases of Urinary Tract Infections (UTIs). The aim of the present study is to isolate UPEC from urine samples from UTI patients, to evaluate the virulence factors and antimicrobial susceptibility pattern.

Material and methods: A total of 100 E.coli isolates out of 292 urine samples collected from clinically suspected UTI were taken for the study and detection of virulence factors such as haemolysin production, haemagglutination and cell surface hydrophobicity were done. Antibiotic susceptibility test was done by Kirby Baur disc diffusion method as per the CLSI guidelines.

Results: The incidence of UTI was more in females 52% and haemolysis was seen in 32%, haemagglutination in 40%, and cell surface hydrophobicity in 24% of strains. The UPEC strains were more resistant to Ampicillin (52%) followed by Cotrimoxazole (46%), Norfloxacin (42%) and Ceftriaxone (40%). Most of the strains were sensitive to Meropenem (80%) followed by Amikacin (78%) and Nitrofurantoin (72%).

Conclusion: As there is a significant association between the virulence and antimicrobial resistance of UPEC, a routine testing of these

factors is recommended and further studies at molecular level are necessary.

Key words: Urinary tract infections, uropathogenic Escherichia coli, virulence factors, antimicrobial susceptibility testing, multidrug resistance

INTRODUCTION

UTIs are the most commonly encountered bacterial infections both in community and health care settings¹. Common UTI causing bacterial pathogens belong to family Enterobacteriaceae, Escherichia coli being the most common member isolated. It has been known that certain serotypes of E.coli are consistently associated with uropathogenicity and are designated as uropathogenic E.coli that expresses chromosomally encoded virulence markers². The common virulence factors include surface hydrophobicity, colonization factor, capsule, serum resistance, resistance to phagocytosis, haemolysin, enterotoxin and siderophore, fimbriae and haemagglutination³.

Uropathogenic strains account for 90% of all UTIs among ambulatory patients and upto 50% of all nosocomial UTIs⁴. The virulence of individual strains in a given infection is determined by the presence and actual expression of the virulence genes present in them and also by the

environmental conditions in the host⁵. The markers of UPEC are expressed with different frequencies in different disease states ranging from asymptomatic bacteriuria to chronic pyelonephritis⁶. Considering the high degree of morbidity and mortality due to UTIs caused by UPEC, the present study was conducted in a tertiary care hospital in Visakhapatnam to study the virulence factors and the antibiotic sensitivity pattern of the UPEC strains isolated from the urine samples of patients suffering from urinary tract infections.

MATERIAL AND METHODS:

This prospective study was conducted in the Department of Microbiology at a tertiary care hospital in Visakhapatnam in 2019. Patients of age group 18 and above were included and paediatric age group was excluded from the study. A total of 100 E.coli strains isolated from 292 urine samples were taken for the study. The samples were processed immediately as per the standard guidelines in the lab. The isolates were taken for the detection of virulence factors and antibiotic susceptibility testing.

Haemolysin production:

The isolates were inoculated onto 5 %sheep blood agar and incubated overnight at 37 degree Celsius and observed for a zone of complete lysis around the colony (Fig.1). *E.coli* ATCC 25922 was used as a negative control.



Figure 1 – Haemolysis

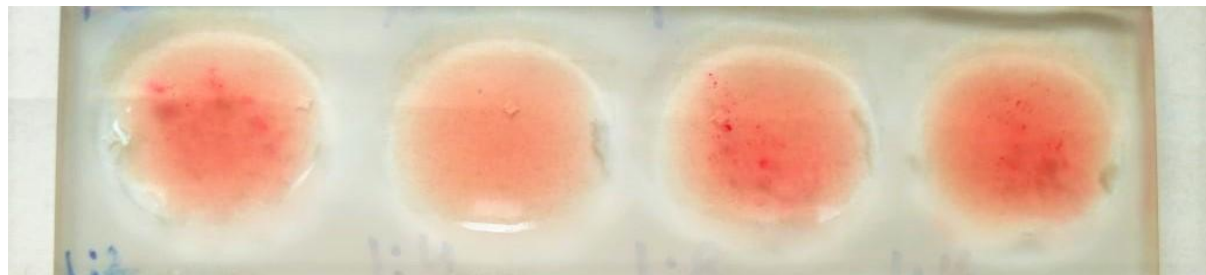
Haemagglutination:

5 ml of group A positive venous blood was collected and washed 3 times in physiological saline and a 3% suspension in fresh saline was made

A drop of very dense bacillary deposit was mixed with an equal drop of red cell suspension on a VDRL slide and it was rocked gently for 5 min. The haemagglutination produced was seen with naked eye as coarse clumps and microscopically. A drop of 2% solution of D-mannose (specifically inhibits type 1 fimbrial haemagglutination) was then added.

Observation: If haemagglutination occurs in the presence of D-mannose, it is considered as mannose resistant (MRHA) (fig.2). If haemagglutination is inhibited by D-mannose, it will be mannose sensitive haemagglutination (MSHA) (fig.3).

MRHA



Positive control

Negative control

Test

D-Mannose

Figure 2

MSHA

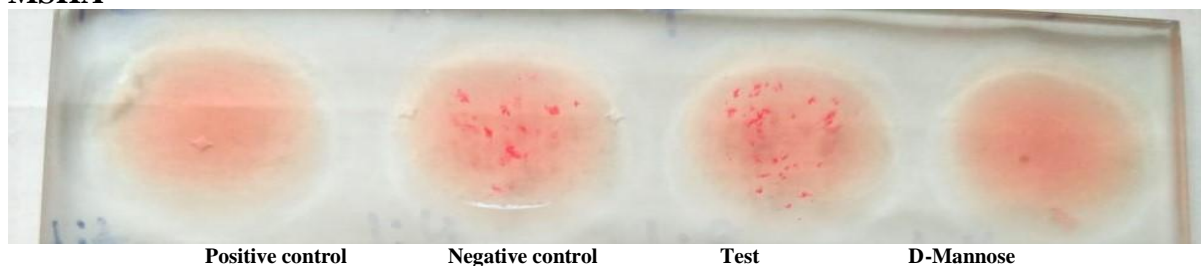


Figure 3

E. coli ATCC 25922 were used as a positive control for mannose sensitive haemagglutination.

Cell Surface Hydrophobicity: Different molar concentrations of ammonium sulphate – 1M, 1.4M, 2M was prepared.

40µl of PBS (Phosphate Buffer solution) was taken in the first well of the VDRL slide. 40µl of 1M, 1.4M, 2M concentrations of ammonium sulphate were

taken in each of the other wells of VDRL slide. 40µl of *E. coli* suspension was added to each well. The clumps found in different molar concentrations of ammonium sulphate observed under naked eye. Strains were considered hydrophobic if they aggregated with 1.4M concentration of ammonium sulphate. *E. coli* ATCC 25922 was used as a negative control for cell surface hydrophobicity (fig 4 and fig 5).

CSH Test Negative

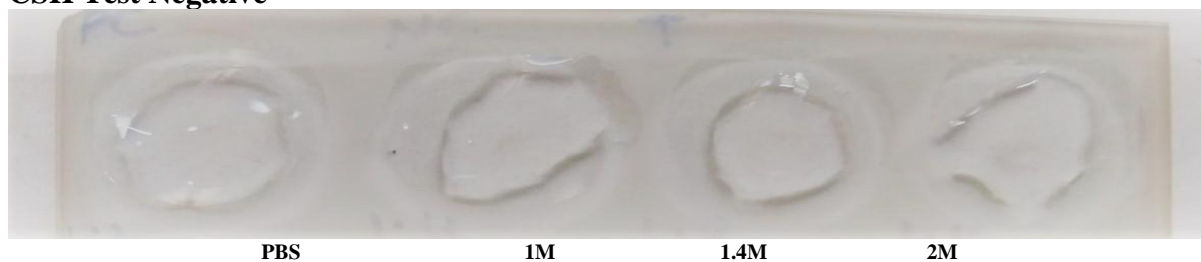


Figure 4

CSH Test Positive

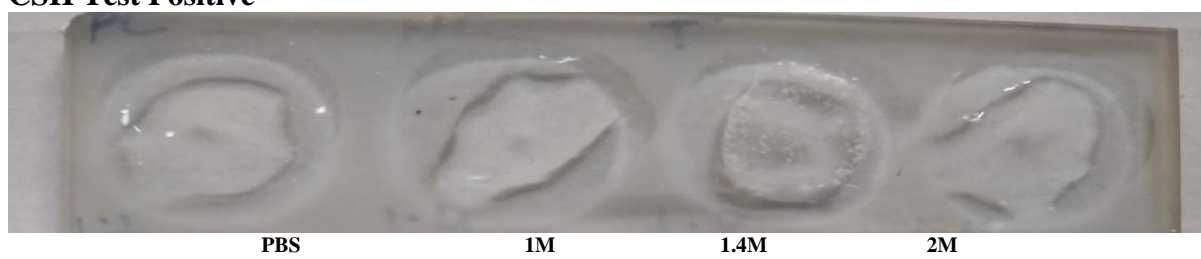


Figure 5

Antimicrobial susceptibility:

Antimicrobial susceptibility testing was done on Mueller Hinton Agar by Kirby Bauer disc diffusion method as per CLSI guidelines.

RESULTS

A total of 292 urine samples received from symptomatic cases of urinary

tract infection with significant bacteriuria were processed. Out of these, 100 *E. coli* isolates were obtained and studied for the virulence factors and antibiotic susceptibility pattern.

Out of the 100 *E. coli* isolated samples, 52% were from females and 48% were from males. The incidence of UPEC was more in the age group of 20-39 years.

Among 100 isolates, 32% showed haemolysis, 40% showed mannose resistant haemagglutination, 4% showed mannose sensitive haemagglutination and 24% showed cell surface hydrophobicity.

The UPEC strains were more resistant to Ampicillin (52%) followed by

Cotrimoxazole (46%), Norfloxacin (42%) and Ceftriaxone (40%). Most of the strains were sensitive to Meropenem (80%) followed by Amikacin (78%) and Nitrofurantoin (72%) (Table 1).

Table 1

CORRELATION BETWEEN ANTIMICROBIAL RESISTANCE AND VIRULENCE FACTORS OF UPEC							
VIRULENCE FACTOR	AK	AMP	CTR	COT	MRP	NIT	NX
HEMOLYSIN							
Positive (n=32)	12 (37.5%)	22 (68.7%)	20 (62.5%)	24 (85.7%)	10 (31.2%)	14 (43.7%)	22 (68.7%)
Negative (n=68)	10 (14.7%)	30 (44.1%)	20 (29.4%)	22 (32.3%)	10 (14.7%)	14 (20.5%)	20 (29.4%)
HAEMAGGLUTININ							
MRHA positive (n=40)	10 (25%)	32 (80%)	24 (60%)	18 (45%)	12 (30%)	16 (40%)	22 (55%)
MRHA negative (n=60)	12 (20%)	20 (33.3%)	16 (26.6%)	28 (46.6%)	8 (13.3%)	12 (20%)	20 (33.3%)
MSHA positive (n=4)	4 (100%)	4 (100%)	2 (50%)	4 (100%)	-	2 (50%)	4 (100%)
MSHA negative (n=96)	18 (18.7%)	48 (50%)	38 (39.5%)	42 (43.7%)	20 (20.8%)	26 (27%)	38 (39.5%)
CELL SURFACE HYDROPHOBICITY							
CSH positive (n=24)	12 (50%)	16 (66.6%)	8 (33.3%)	18 (75%)	8 (33.3%)	12 (50%)	14 (58.3%)
CSH negative (n=76)	10 (13.1%)	36 (47.3%)	32 (42.1%)	28 (36.8%)	12 (15.7%)	16 (21%)	28 (36.8%)

DISCUSSION

Considering the high degree of morbidity and mortality of UTIs, the subject of UPEC is receiving increasing attention. UTIs which are not properly treated from their onset can become a renal threat in time, finally leading to renal failure. In general, the more virulence factors a strain expresses, the more severe an infection it is able to cause⁷. The occurrence of multiple virulence factors in UPEC strains further strengthens the concept of association of UPEC with urinary pathogenicity⁸. These virulence factors enable some members of the normal flora to elicit an infection by overcoming the host defense mechanisms. Virulence factors enable E.coli to colonize selectively the mucosal uro-epithelium, evoke an inflammatory reaction and eventually proceed from lower urinary tract to renal cavities and tissue invasion. The capacity of E.coli to produce many virulence factors contribute to its pathogenicity⁹.

UTI is the most repeatedly diagnosed kidney and urologic disease and E.coli is by far the most common etiologic

agent¹⁰. UTI is the second most common cause of bacterial infection in humans and thus represents a major source of human discomfort². E.coli strains causing UTIs typically agglutinate human erythrocytes despite the presence of mannose and this was mediated by fimbriae⁸. The ability of E.coli to adhere to the uro-epithelium is mediated by fimbriae, thereby resisting elimination by the flow of urine. Adhesion therefore is an important step in the pathogenesis of UTI². UPEC strain take advantage of a variety of virulence properties in order to colonize and establish an UTI. Bacterial adherence and colonization of UPEC are mediated by the expression of several types of fimbrial and non-fimbrial adhesions. Haemolysin is produced by many UPEC, which may be involved in kidney disease¹¹.

In the present study, 52% of samples were from females which correlate with Sanjay Singh Kaira et al¹² who reported 56.09%, Mittal et al¹³ (53.3%) and Chhaya et al¹⁴ (53%).

The incidence of UPEC was more in the age group of 20-39 years in the present

study, which correlates with Hooton et al¹⁵, Kabugo et al¹⁶ and Chhaya et al¹⁴.

Haemolysin production was observed in 32% of isolates in our study which correlates with Chhaya et al¹⁴ (32.3%), and Nazish Fatima et al¹⁷ (30%) whereas Karam et al¹⁸ reported higher incidence of 44.5% and Mittal et al¹³ 47%. Kauser et al¹⁹ reported lower incidence of 21% and Sharma et al²⁰ 25%.

In our study, MRHA and MSHA were observed in 40% and 4% which correlates with Kaira et al¹², who reported 41% and 5%, whereas Chhaya et al¹⁴ reported 52.3% and 8.5%.

Cell surface hydrophobicity was observed in 24% of isolates in our study which correlates with Raksha et al⁸ who reported 26.36% and Kaira et al¹² who reported 27.64% whereas Dorota et al²¹ reported 74% and Mittal et al¹³ reported 61%.

The antibiotic susceptibility pattern observed in our study correlates with Tabasi et al²², Karam et al¹⁸ and Chhaya et al¹⁴.

CONCLUSION

Multidrug resistance was observed at a higher rate among UPEC strains. Therefore, a significant association has been found between the Virulence Factors of UPEC and antimicrobial resistance in UPEC. A routine testing of these factors and co-relation with AMR is thus recommended. Since most urovirulent strains express multiple virulent factors simultaneously, further studies at molecular level are necessary.

REFERENCES

1. Ranjan KP, Ranjan N. Complicated urinary tract infection caused by extended spectrum β -lactamase-producing Escherichia coli. Urol Ann 2014;6:112-3.
2. Vagarali MA, Karadesai SG, Patil CS, Metgud SC, Mutnal MB. Haemagglutination and Siderophore production as the urovirulence markers of Uropathogenic Escherichia coli. Indian J Med Microbiol. 2008;26(1):68-70
3. Soto SM, Smithson A, Martinez JA, Horcajada JP, Mensa J, Vila J, et al. Biofilm formation in uropathogenic Escherichia coli strains: Relationship with prostatitis, urovirulence factors and antimicrobial resistance. J Urol 2007;177:365-8.
4. Steadman R, Topley N (1998) The virulence of Escherichia coli in urinary tract, In: Urinary tract infections. Chapman and Hall publication, London.
5. Sharma S, Bhat G K, Shenoy S. Virulence factors and drug resistance in Escherichia coli isolated from extraintestinal infections. Indian J Med Microbiol 2007; 25(4):369-373.
6. Raksha R, Srinivasa H, Macaden R S. Occurrence and characterization uropathogenic Escherichia coli in urinary tract infections. Indian J Med Microbiol 2003; 21(2):102-107.
7. Mandal P, Kapil A, Goswami K, Das B, Dwivedi SN. Uropathogenic Escherichia coli causing Urinary Tract Infections. Indian J Med Res. 2001;114:207-11.
8. Raksha R, Srinivasa H, Macaden RS. Occurrence & Characterization of Uropathogenic Escherichia coli in UTIs. Indian J Med Microbiol. 2003;21(2):102-7.
9. Biswas D, Gupta P, Prasad R, Singh V, Arya M, Kumar A. Choice of antibiotics for empirical therapy of acute cystitis in a setting of high antimicrobial resistance. Indian J Med Sci. 2006;60(2):53-8.
10. Kao JS, Stuckes DM, Warren JW, Mobley H.L. Pathogenicity island sequences of pyelonephritogenic Escherichia coli CFT073 are associated with virulent Uropathogenic strains. Infect Immuno. 1997;65(7):2812-20.
11. Georgi S, Pisareva E, Markova N. Virulence of uropathogenic Escherichia coli. J Culture Collections. 2008;6:3-9.
12. Kaira SS, Pai C. Study of uropathogenic Escherichia coli with special reference to its virulence factors. Int J Community Med Public Health 2018;5:177-81.
13. Mittal S, Sharma M, Chaudhary U. Study of virulence factors of uropathogenic Escherichia coli & its antibiotics susceptibility pattern. Indian J Pathol Microbiol. 2014;57(1):61-4
14. Shah Chhaya, Baral R, Bartaula B, Shrestha LB. Virulence factors of uropathogenic Escherichia coli (UPEC) and correlation with antimicrobial resistance. BMC

- Microbiol. 2019 Sep 2;19(1):204. doi: 10.1186/s12866-019-1587-3.
15. Hooton TM, Scholes D, Hughes JP, Winter C, Roberts PL, Stapleton AE, et al. A prospective study of risk factors for symptomatic urinary tract infections in young women. *Ne Eng J Med* 1996;335: 468-74.
 16. Kabugo D, Kizito S, Ashok DD, Graham KA, Nabimba R, Namunana S, Kabaka MR, Achan B, Najjuka FC. Factors associated with community-acquired urinary tract infections among adults attending assessment centre, Mulago Hospital Uganda. *Afr Health Sci.* 2016 Dec; 16(4):1131-1142. doi: 10.4314/ahs.v16i4.31
 17. Fatima N, Agrawal M, Shukla I, Khan PA (2012) Characterization of Uropathogenic E. coli in relation to virulence Factors. 1:342. doi:10.4172/scientificreports.342
 18. Karam MRA, Habibi M, Bouzari S. Relationships between Virulence Factors and Antimicrobial Resistance among *Escherichia coli* Isolated from Urinary Tract Infections and Commensal Isolates in Tehran, Iran. *Osong Public Health Res Perspect.* 2018 Oct;9(5):217-224. doi: 10.24171/j.phrp.2018.9.5.02.
 19. Kauser Y, Chunchanur SK, Nadagir SD, Halesh LH, Chandrashekhar MR. Virulence factors, serotypes & antimicrobial susceptibility of *Escherichia coli* in urinary tract infection. *Al Amen J Med Sci.* 2009;2:47-1
 20. Sharma S, Bhat GK, Shenoy S. Virulence factors and drug resistance in *Escherichia coli* isolated from extraintestinal infections. *Indian J Med Microbiol.* 2007;25(4):369-73
 21. Dorota W. Virulence factors of Uropathogenic *Escherichia coli* strains isolated from children with chronic pyelonephritis. *Adv Clin Exp Med.* 2007; 16(5):651-7.
 22. Tabasi M, Asadi Karam MR, Habibi M, Yekaninejad MS, Bouzari S. Phenotypic Assays to Determine Virulence Factors of Uropathogenic *Escherichia coli* (UPEC) Isolates and their Correlation with Antibiotic Resistance Pattern. *Osong Public Health Res Perspect.* 2015;6(4):261-268. doi:10.1016/j.phrp.2015.08.002
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